



Product Sheet

Autobioluminescent HEK293 Cells

Handling Upon Receipt

Inspect packaging for damage or leaks. Remove the vial containing cells from dry ice and store in liquid nitrogen vapor (< -130 °C) or immediately thaw and culture.

Warranty

This product is warranted viable for 30 days from the date of shipment and is valid only if the product is stored and cultured according to the information provided on this sheet. The use of medium components other than those recommended by 490 BioTech on this sheet will invalidate this warranty.

Disclaimers

While 490 BioTech makes every effort to include accurate and up to date information on this product information sheet, 490 BioTech makes no warranties or representations of its accuracy. This product is sent with the condition that you are responsible for its safe storage, handling, and use. 490 BioTech is not liable for any damages or injuries arising from the receipt and/or use of this product.

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Cellular Background

Organism: *Homo sapiens*

Tissue Type: Kidney

Growth Format: Adherent

Recommended Growth Medium

The following medium is recommended for cellular growth:

- DMEM
- 10% FBS
- 1X Sodium Pyruvate
- 1X Antibiotic/Antimycotic
- 100 µg G418/ml

Thawing Frozen Cells

1. Incubate the vial in a 37 °C water bath with gentle agitation until contents have thawed (thawing should occur in approximately 2 minutes). To reduce the possibility of contamination, do not submerge the O-ring or cap during the thawing process.
2. Spray the thawed vial with 70% ethanol and transfer to a sterilized environment. All remaining steps should be performed using aseptic technique.
3. Transfer the full contents of the vial to a 15 ml centrifuge tube containing 9 ml of complete growth medium pre-incubated to 37 °C and centrifuge at 125 x g for 7 minutes.
4. Resuspend the cell pellet in complete growth medium pre-incubated to 37 °C and transfer to an appropriate cell culture vessel. 25 cm² or 75 cm² culture flasks are recommended for initial thawing procedures.
5. Incubate the cells at 37 °C 5% CO₂ in a humidity controlled environment and monitor for growth.

Routine Growth and Maintenance

1. Remove and discard spent culture medium.
2. Rinse cells with an appropriate volume of sterile, 37 °C PBS.
3. Add an appropriate volume of Trypsin-EDTA solution to the flask and incubate until cells have detached (detachment usually occurs within 2 to 15 minutes depending on if the incubation is performed at room temperature or at 37 °C).
4. Resuspend the detached cells in an appropriate amount of pre-incubated 37 °C complete growth medium.
5. Aliquot cells into new culture vessels containing pre-incubated 37 °C complete growth medium.
6. Incubate the cells at 37 °C 5% CO₂ in a humidity controlled environment.

Subcultivation ratio: It is recommended that cells be subcultured at a ratio between 1:3 and 1:10 as needed.

Medium Refreshment: It is recommended that medium be refreshed every 2 to 3 days as needed.

Biosafety Information

These cells are treated as biosafety level 2. Appropriate safety procedures should always be followed when working with or using these cells. More information can be obtained from the US Department of Health and Human Services Centers for Disease Control and Prevention and National Institutes of Health's *Biosafety in Microbiological and Biomedical Laboratories*.